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Genome-wide association study of conformation and milk yield in mixed-breed dairy goats

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ABSTRACT

Identification of genetic markers that affect economically important traits is of high value from a biological point of view, enabling the targeting of candidate genes and providing practical benefits for the industry such as wide-scale genomic selection. This study is one of the first to investigate the genetic background of economically important traits in dairy goats using the caprine 50K single nucleotide polymorphism (SNP) chip. The aim of the project was to perform a genome-wide association study for milk yield and conformation of udder, teat, and feet and legs. A total of 137,235 milk yield records on 4,563 goats each scored for 10 conformation traits were available. Out of these, 2,381 goats were genotyped with the Illumina Caprine 50K BeadChip (Illumina Inc., San Diego, CA). A range of pseudo-phenotypes were used including deregressed breeding values and pseudo-estimated breeding values. Genome-wide association studies were performed using the multi-locus mixed model (MLMM) algorithm implemented in SNP & Variation Suite v7.7.8 (Golden Helix Inc., Bozeman, MT). A genome-wide significant [$-\log_{10}(P\text{-value}) > 5.95$] SNP for milk yield was identified on chromosome 19, with additional chromosome-wide significant ($-\log_{10}(P\text{-value}) > 4.46$) SNP on chromosomes 4, 8, 14, and 29. Three genome-wide significant SNP for conformation of udder attachment, udder depth, and front legs were identified on chromosome 19, and chromosome-wide SNP were found on chromosomes 4, 5, 6, 10, 11, 12, 13, 14, 15, 16, 17, 18, 21, 23, and 27. The proportion of variance explained by the significant SNP was between 0.4 and 7.0% for milk yield and between 0.1 and 13.8% for conformation traits. This study is the first attempt to identify SNP associated

with milk yield and conformation in dairy goats. Two genome-wide significant SNP for milk yield and 3 SNP for conformation of udder attachment, udder depth, and front legs were found. Our results suggest that conformation traits have a polygenic background because, for most of them, we did not identify any quantitative trait loci with major effect.

Key words: genome-wide association study (GWAS), dairy goat, conformation, quantitative trait loci (QTL)

INTRODUCTION

Conventional goat breeding schemes are often hampered by the cost of measuring phenotypes and maintaining accurate pedigree and data recording. Many goat breeders do not perform routine data recording or do so on a very basic level. Identification of genetic markers and candidate genes that affect economically important traits is therefore of high value from a biological point of view, as well as providing practical benefits to industry. Compared with that in cattle and sheep, the number of QTL studies in goats is limited; however, at present, goat QTL are not included in Animal QTLdb database (<http://www.animalgenome.org/cgi-bin/QTLdb/index>), which makes it difficult to judge the exact number of studies in this species (for review, see Amills, 2014).

Quantitative trait loci for traits such as birth and weaning weight (Mohammad Abadi et al., 2009; Visser et al., 2013), hair fiber characteristics (Cano et al., 2007; Visser et al., 2011), growth (Mohammad Abadi et al., 2009), body conformation (Marrube et al., 2007), parasite resistance (Bolormaa et al., 2010; De La Chevrotière et al., 2012), milk traits (Roldán et al., 2008), and α_{S1} -casein (Sacchi et al., 2005; Hayes et al., 2006; Dagnachew et al., 2011) have been identified in goats using microsatellite markers. With the availability of the Illumina Caprine 50K BeadChip (Illumina Inc., San Diego, CA; Tosser-Klopp et al., 2012, 2014), it is now possible to perform genome-wide scans in goats

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with much greater resolution. To date, the chip has been utilized to find QTL associated with milk and type traits (Maroteau et al., 2013; Martin et al., 2017), polledness (Kijas et al., 2013), milking speed (Palhière et al., 2014), coat color (Becker et al., 2015; Martin et al., 2016a), wattles (Reber et al., 2015), and supernumerary teats (Martin et al., 2016b). In addition, due to the application of sequencing and whole-genome optical mapping, information on gene annotation for goats has become available (Dong et al., 2013). The goat genome assembly has been refined and updated using dense radiation hybrid mapping (Du et al., 2014).

In cattle, feet and leg conformation has been genetically linked to claw disorders (van der Waaij et al., 2005), although there is evidence that animals with good udder conformation are less sensitive to high yields than those with poor udder conformation (Philipsson and Lindhé, 2003). In dairy goats, there is evidence of genetic correlations with production traits such as milk yield (McLaren et al., 2016) and productive life, as well as functional productive life (Castañeda-Bustos et al., 2017). Udder and teat conformation have also been linked with elevated SCC and clinical mastitis (Rupp and Boichard, 1999; Rupp et al., 2011; Huntley et al., 2012) as well as milking speed (Sewalem et al., 2011). Conformation traits are therefore important from both a health and welfare perspective and a production perspective; however, these traits are costly to phenotype. Identifying SNP associated with QTL that significantly affect a trait would allow for marker-assisted selection, saving time and money on phenotyping adult animals. The main objective of this study was to perform genome-wide association studies (**GWAS**) to identify SNP associated with conformation of udder, teat, legs and feet, as well as milk yield. This study is one of the first to investigate genetic background of economically important traits in dairy goats using the caprine 50K SNP chip.

MATERIALS AND METHODS

Data

A total of 5,026 first-lactation goats were scored for 10 conformation traits. Goats were a composite breed of Saanen, Toggenburg, and Alpine. A more detailed description of breed composition investigated with a principal component analysis is given in Mucha et al. (2015). Scoring was performed on 2 farms owned by the same farming business, over 2 yr (2013 and 2014) by the same scorer. All goats in their first lactation were scored once on both participating farms. Animals were ranked on a linear scale (1–9), similar to the scoring

system developed by the French dairy goat breeders' association (CAPGENES) and used by Manfredi et al. (2001) and Rupp et al. (2011). A detailed description of the scoring system can be found in McLaren et al. (2016). The data contained information for udder traits (udder furrow, udder depth, udder attachment), teat traits (teat shape, teat angle, teat placement), and traits relating to legs and feet (front legs, back legs, front feet, back feet). A total of 137,235 milk yield records were available for 4,563 of the scored animals. The pedigree contained 34,391 individuals over 15 generations, with a total of 296 and 12,468 registered sires and dams, respectively.

Genotyping

Initially, 1,960 animals were selected for genotyping to build a reference population for implementation of genomic selection. All of the available sires (150 individuals) were sampled, and this set was subsequently supplemented by females. Details on the structure of the genotyped population and selection method are given in Mucha et al. (2015). Subsequently, all female animals selected as replacements and all males used in the population were genotyped. In total, 2,381 goats with milk yield data and 402 of the conformation-scored goats were genotyped. Selection of animals for genotyping was done purely based on milk production, because this was the main breeding objective on the participating farms. That is why only 402 of the genotyped goats had conformation data. Animals were genotyped commercially with the Illumina Caprine 50K BeadChip (Illumina Inc.; Tosser-Klopp et al., 2012) at Edinburgh Genomics (Edinburgh, UK). Single nucleotide polymorphisms were removed from further analyses if they were not in Hardy-Weinberg equilibrium, had minor allele frequency <0.05, were monomorphic, had a call rate of <0.95, or if the Illumina GenCall (GC score) was below 0.6. Missing genotypes (0.2% of all genotyped SNP) were imputed as homozygous for the major allele. Three animals were removed because they had call rate <0.90. This resulted in a data set with 2,378 goats with milk yield data and 402 conformation-scored goats genotyped for 44,798 SNP from 29 chromosomes (the X chromosome was excluded from the analysis).

Calculation of Deregressed Breeding Values

Basic statistics describing the data were estimated using R (R Development Core Team, 2011). Appropriate statistical models for conformation traits and milk yield were developed in previous studies on the same population (Mucha et al., 2014; McLaren et al., 2016).

Conformation Traits. Breeding values for conformation traits were predicted using records from 5,026 animals using the software package MIX99 (Lidauer et al., 2011) with the following model:

$$\mathbf{y} = \mathbf{X}\mathbf{b} + \mathbf{Z}\mathbf{a} + \mathbf{e}, \quad [\text{model 1}]$$

where \mathbf{y} is the vector of observations for one of the conformation traits; \mathbf{b} is the vector of fixed effects consisting of lactation stage (100-d periods from 1 to 700 DIM), farm (2 levels), birth year (2006 to 2013), and scoring year (2013 and 2014); \mathbf{a} is the vector of random animal effects; \mathbf{e} is the vector of random residuals; and \mathbf{X} and \mathbf{Z} are incidence matrices relating records to their respective effects.

Breeding values were predicted in a multi-trait analysis with the 10 conformation traits. Random effects were assumed normally distributed with zero means and the following covariance structure:

$$\text{Var} \begin{bmatrix} \mathbf{a}_1 \\ \mathbf{a}_2 \\ \vdots \\ \mathbf{a}_n \\ \mathbf{e}_1 \\ \mathbf{e}_2 \\ \vdots \\ \mathbf{e}_n \end{bmatrix} = \begin{bmatrix} \mathbf{A}\sigma_{a1}^2 & \mathbf{A}\sigma_{a12} & \dots & \mathbf{A}\sigma_{a1n} & 0 & 0 & 0 & 0 \\ & \mathbf{A}\sigma_{a2}^2 & \dots & \mathbf{A}\sigma_{a2n} & 0 & 0 & 0 & 0 \\ & & \dots & \vdots & \vdots & \vdots & \vdots & \vdots \\ & & & \mathbf{A}\sigma_{an}^2 & 0 & 0 & 0 & 0 \\ & & & & \mathbf{I}\sigma_{e1}^2 & \mathbf{I}\sigma_{e12} & \dots & \mathbf{I}\sigma_{e1n} \\ & & & & & \mathbf{I}\sigma_{e2}^2 & \dots & \mathbf{I}\sigma_{e2n} \\ & & & & & & \dots & \vdots \\ & & & & & & & \mathbf{I}\sigma_{en}^2 \end{bmatrix},$$

symm

where \mathbf{A} is the pedigree-based relationship matrix, \mathbf{I} are identity matrices, σ_a^2 and σ_e^2 are the genetic and residual variances, and indices 1 to n indicate the 10 conformation traits.

Milk Yield. Breeding values for milk yield were predicted with the following random regression model (Mucha et al., 2014):

$$\mathbf{y} = \mathbf{X}\mathbf{b} + \mathbf{Z}\mathbf{a} + \mathbf{W}\mathbf{p} + \mathbf{e}, \quad [\text{model 2}]$$

where \mathbf{y} is the vector of test-day milk yield observations; \mathbf{b} the vector of fixed effects consisting of herd test-day, age at kidding, year-season, and fixed lactation curves modeled by fitting Legendre polynomials (Kirkpatrick et al., 1990) of fourth order; \mathbf{a} is a 1×3 vector of random regression coefficients (Legendre polynomials of second order) for the animal effect; \mathbf{p} is the 1×3 vector of random regression coefficients (Legendre polynomials of second order) for the permanent environment effect; and \mathbf{e} is the vector of random residual effect. The matrix \mathbf{X} is the incidence matrix

for fixed effects, and \mathbf{Z} and \mathbf{W} are matrices of Legendre polynomials of DIM of second order for random animal and permanent environment effect, respectively.

Random effects were assumed normally distributed with zero means and the following covariance structure:

$$\text{Var} \begin{bmatrix} \mathbf{a} \\ \mathbf{p} \\ \mathbf{e} \end{bmatrix} = \begin{bmatrix} \mathbf{A} \otimes \mathbf{G} & 0 & 0 \\ & \mathbf{I} \otimes \mathbf{P} & 0 \\ \text{symm} & & \mathbf{I}\sigma_e^2 \end{bmatrix},$$

where \mathbf{G} and \mathbf{P} are 3×3 (co)variance matrices of the random regression coefficients for the animal and permanent environment effects, respectively.

The software package MIX99 was also used for deregression, using a full animal pedigree with effective offspring contributions (*EOC*) as weighting factors for each of the conformation traits and milk yield. The *EOC* were calculated for each trait as

$$EOC_{ij} = \frac{rel_{ij} \cdot kdau_i}{1 - rel_{ij}}, \text{ and } kdau_i = \frac{4 - h_i^2}{h_i^2},$$

where rel_{ij} is the reliability of EBV for trait i of animal j , and h_i^2 is the heritability of trait i .

Deregressed EBV were used as pseudo-phenotypes in the subsequent GWAS analysis.

Calculation of Pseudo-EBV

As an alternative to deregressed EBV, a second alternative phenotype for the conformation scores and milk yield was considered. The same models (model 1 and 2) were fitted for conformation scores and milk yield, but with an identity matrix \mathbf{I} fitted instead of the pedigree-based relationship matrix \mathbf{A} . This was done to remove any influence of pedigree relationships (contributions of parents and relatives) on an animal's own breeding value. These models were fitted to test whether deregressed EBV lead to false-positive results in GWAS.

Genome-Wide Association Analysis

The GWAS was performed using the multi-locus mixed model (MLMM) algorithm (Segura et al., 2012) implemented in SNP & Variation Suite v7.7.8 (Golden Helix Inc., Bozeman, MT). An additive genetic model was used, where the major homozygous genotype was coded as 0, heterozygous as 1, and minor homozygous as 2. The position of the markers was determined using the map CHIR_1.0 (Dong et al., 2013). The following

model with random polygenic effect, and the genotypes at single SNP markers as fixed effects was used:

$$\mathbf{y} = \mathbf{X}\boldsymbol{\beta} + \mathbf{Z}\mathbf{a} + \mathbf{e}, \quad [\text{model 3}]$$

where \mathbf{y} is the vector of phenotypes for the analyzed trait; $\boldsymbol{\beta}$ is a vector of coefficients for the SNP effects and fixed effects (as defined for model 1, only applicable for raw conformation scores – phenotype (2) defined below); \mathbf{a} is the vector of random animal effects; \mathbf{e} is the vector of random residual effects; and \mathbf{X} and \mathbf{Z} are incidence matrices relating observations to fixed and random animal effects, respectively. Random effects were assumed normally distributed with zero means and the following covariance structure:

$$\text{Var} \begin{bmatrix} \mathbf{a} \\ \mathbf{e} \end{bmatrix} = \begin{bmatrix} \mathbf{G}\sigma_a^2 & 0 \\ 0 & \mathbf{I}\sigma_e^2 \end{bmatrix},$$

where \mathbf{G} is the genomic relationship matrix (VanRaden, 2008) calculated as

$$\mathbf{G} = \frac{\mathbf{SS}'}{2 \sum_{i=1}^N p_i (1 - p_i)},$$

where \mathbf{S} is a centered incidence matrix of SNP genotypes, N is the number of SNP markers, and p_i is allele frequency of marker i .

Genome-wide association analyses were performed using the following phenotypes: (1) deregressed EBV for conformation scores and milk yield; (2) raw conformation scores; (3) pseudo-EBV (**psEBV**) for conformation scores (only animals that were scored) and milk yield (only animals with own milk records); and (4) filtered deregressed EBV for conformation scores, which was a subset of data used in phenotype 1, where only animals with their own conformation scores or reliability of their EBV > 0.6 were included in GWAS. Deregressed EBV were chosen as a phenotype because they can maximize the use of information available on each animal (repeated records per animal), as well as progeny information in the case of sires. However, for conformation traits that have lower heritabilities and fewer phenotyped animals, the reliability of the deregressed EBV was relatively low. Therefore, to avoid spurious results, additional phenotypes (2–4) were included in the analysis.

Quantile-quantile (**Q-Q**) plots were used to analyze the extent to which the observed distribution of the test statistic followed the expected (null) distribution.

This was done to assess potential systematic bias due to population structure or analytical approach. Bonferroni correction was applied to obtain significance thresholds. A SNP was significant at the genome-wise level when the $-\log_{10}(P\text{-value})$ was greater than $-\log_{10}(0.05/N)$, where N is the total number of markers, which was equal to 44,798 and gave us a genome-wise threshold of 5.95. Chromosome-wise significant SNP had an associated $-\log_{10}(P\text{-value})$ above $-\log_{10}(0.05/n)$, where n is the number of markers on a given chromosome. The proportion of variance explained by SNP (*pve*) was calculated as

$$pve = \frac{mrss_{h0} - mrss_k}{mrss_{h0}},$$

where $mrss_{h0}$ is the Mahalanobis root sum of squares (*mrss*) for the null hypothesis and $mrss_k$ is the same for marker k .

RESULTS

Phenotype Statistics

Descriptive statistics for milk yield and the raw conformation scores of the animals used in this study are presented in Table 1. Mean test-day milk yield in the genotyped group of 2,165 goats was 3.89 with a standard deviation (**SD**) of 1.73. For a review of more detailed statistics on milk yield in the analyzed population, see Mucha et al. (2014). In total, 402 of the genotyped animals had conformation data available. Front legs and back legs had the smallest SD of 0.38 and 0.57, respectively. This reflects the fact that majority of the animals had a score of 5 for the 2 traits. The remaining traits displayed larger variation but their distributions were often skewed toward higher scores. Deregressed EBV for conformation traits were estimated for all 2,378 genotyped goats. However, due to low mean reliability of the EBV, which were between 0.15 and 0.25, these traits were considered not informative (Table 2). Higher reliability was obtained with a subset of that data for animals with conformation records or high reliability EBV (phenotype 4, as described above). In this case, the data set contained records of 417 to 477 animals (depending on the trait) with a mean reliability of between 0.24 and 0.52 (Supplemental Table S1; <https://doi.org/10.3168/jds.2017-12919>). The data set was reduced further in case of psEBV, which were estimated only for the 402 conformation-scored goats. The mean of psEBV for all of the analyzed conformation traits was close to 0, with the minimum and maximum values between -1.29 and 1.37 , respectively

Table 1. Number of records (N) and descriptive statistics for raw conformation scores and milk yield (genotyped animals only)

Trait	N	Mean	SD	Minimum	Maximum	SEM
Milk yield	137,235	3.89	1.73	0.2	12.1	<0.01
Udder furrow	397	5.95	1.17	2	8	0.06
Udder depth	402	5.86	1.25	3	9	0.06
Udder attachment	402	7.75	0.77	5	9	0.04
Teat shape	401	4.27	1.14	1	7	0.06
Teat angle	402	4.22	0.83	1	5	0.04
Teat placement	402	3.33	0.85	1	6	0.04
Front legs	402	4.84	0.38	3	5	0.02
Back legs	402	4.69	0.57	2	7	0.03
Front feet	402	8.00	0.89	5	9	0.04
Back feet	402	7.44	0.87	5	9	0.04

(Supplemental Table S2; <https://doi.org/10.3168/jds.2017-12919>). Similar to the raw conformation scores, psEBV had the smallest SD for front legs and back legs (0.03 and 0.09).

Q-Q Plots

The Q-Q plots of expected and observed P -values [$-\log_{10}(P\text{-values})$] obtained from GWAS did not reveal any major population stratification that might have affected the current analysis. The Q-Q plots for the models with deregressed EBV for conformation indicate that there might be false-positive results, as many SNP departed from the expected probability (Supplemental Figure S1; <https://doi.org/10.3168/jds.2017-12919>). This issue did not seem to be present with models based on the remaining phenotypes for conformation (Supplemental Figures S2 and S3; <https://doi.org/10.3168/jds.2017-12919>). The highest departures from the expected probability (in all of the analyzed models) were observed for udder depth, which indicates that there were SNP with highly significant effect on this trait. The Q-Q plots for milk yield also indicated that there were highly significant SNP associated with this trait (Supplemental Figure S1). Values of the inflation factor lambda for the analyzed conformation traits

were highest when deregressed EBV were used as the phenotype, with values between 1.10 and 1.16. Lower lambda coefficients were observed with raw conformation scores, psEBV, and filtered EBV, with lambda between 0.96 and 1.02. Lambda for milk yield was between 0.95 and 0.96 for psEBV and deregressed EBV, respectively. Homogeneity of the analyzed population was also confirmed with a principal component analysis performed in a previous study (Mucha et al., 2015).

Genome-Wide Association Analysis

Profiles of the P -values [presented as $-\log_{10}(P\text{-values})$] for milk yield for all SNP from chromosomes 1 to 29 are presented in Figure 1. Both analyses (with deregressed EBV and psEBV) identified a genome-wide significant SNP on chromosome 19 (Table 3). The SNP snp23995-scaffold244-354162 was located between exons 1 and 2 of the *ALOX12* gene (*GOAT_ENSP00000251535*). A second genome-wide significant SNP for milk yield was identified only with the psEBV model. The SNP snp28093-scaffold300-4036770 was located on chromosome 8, 64,859 bp away from the *SLC1A1* gene (*GOAT_ENSBTAP0000025456*). Additionally, 5 SNP were significant on the chromosome-wise level (Supplemental Table S3; <https://doi.org/>

Table 2. Number of animals (n), descriptive statistics, and reliabilities (Rel) of deregressed EBV for milk yield and conformation scores

Trait	n	Mean	SD	Minimum	Maximum	SEM	Rel
Milk yield	2,378	331.55	188.27	-151.00	1,022.00	3.86	0.94
Udder furrow	2,378	2.05	63.25	-40.55	3,076.80	1.30	0.19
Udder depth	2,378	-0.02	2.26	-11.32	76.77	0.05	0.25
Udder attachment	2,378	-0.23	9.14	-442.92	32.29	0.19	0.20
Teat shape	2,378	-0.67	26.47	-1,285.90	90.85	0.54	0.20
Teat angle	2,378	-4.11	149.2	-7,269.50	69.43	3.06	0.24
Teat placement	2,378	3.38	123.37	-39.90	6,011.80	2.53	0.25
Front legs	2,378	-0.67	20.89	-1,014.00	52.48	0.43	0.14
Back legs	2,378	0.13	5.76	-23.10	279.17	0.12	0.15
Front feet	2,378	2.15	93.10	-3.23	4,538.90	1.91	0.15
Back feet	2,378	4.67	185.49	-67.27	9,041.90	3.80	0.15

Table 3. Genome-wise significant SNP for milk yield and conformation scores

Trait ¹	SNP ²	Model ³	-Log ₁₀ (<i>P</i> -value)	Variance explained (%)	Nearest gene	
					Name ⁴	Distance ⁵ (bp)
MY	8:38821852	3	4.66	0.8	<i>GOAT_ENSBTAP00000025456</i>	64,859
		2	6.1	0.4	<i>(SLC1A1)</i>	
MY	19:26066457	3	38.45	6.0	<i>GOAT_ENSP00000251535</i>	Between exons 1
		2	46.1	7.0	<i>(ALOX12)</i>	and 2
UA	19:26150581	1	3.8	3.5	<i>GOAT_ENSP00000347140</i>	Between exons 6
		2	10.95	10.4	<i>(ASGR2)</i>	and 7
UD	19:26972244	1	13.02	12.5	<i>GOAT_ENSBTAP00000034797</i>	Between exons 14
		2	14.73	13.8	<i>(ALOXE3)</i>	and 15
FL	19:27172738	1	0.45	0.2	<i>GOAT_ENSP00000331086</i>	-5,712
					<i>(ODF4)</i>	
		2	7.31	7.1	<i>GOAT_ENSBTAP00000042124</i>	9,224
					<i>(ARHGEF15)</i>	

¹MY = milk yield; UA = udder attachment; UD = udder depth; FL = front legs.

²Number left of colon represents the chromosome number; number to right specifies the position on the chromosome.

³1 = raw scores; 2 = pseudo EBV; 3 = deregressed EBV.

⁴Gene names with bovine homologs in parentheses.

⁵Positive value denotes the gene located downstream of SNP; negative value denotes the gene located upstream of SNP.

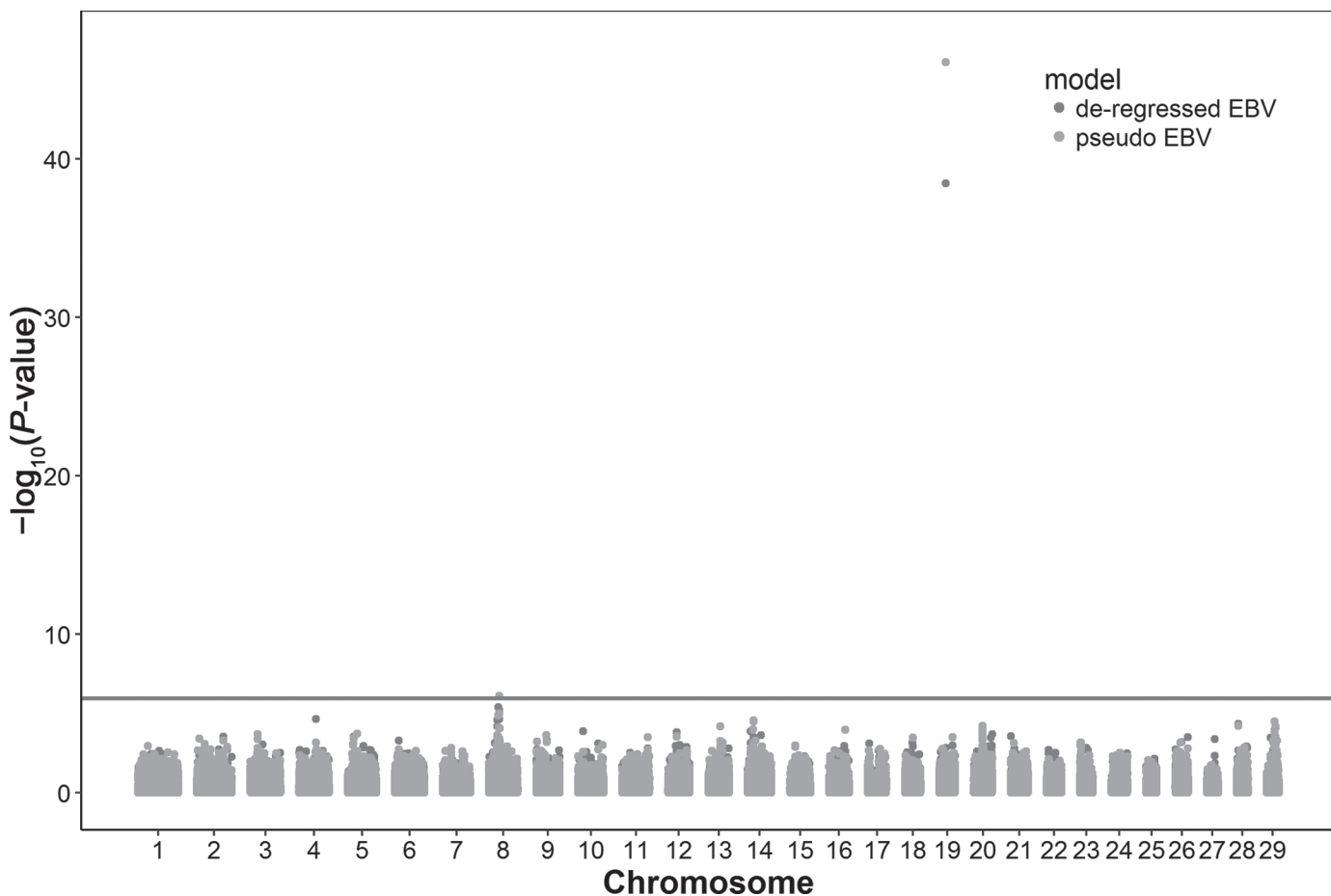


Figure 1. Manhattan plot of genome-wide association study (GWAS) for milk yield using the Illumina Caprine 50K BeadChip (Illumina Inc., San Diego, CA) in dairy goats. *P*-values [$-\log_{10}(P\text{-values})$] for each SNP are shown on the y-axis, and each chromosome number is indicated on the x-axis. The solid (red) line indicates the threshold value for genome-wise significance. Deregressed EBV and pseudo-EBV (calculated by adjusting for fixed effects using BLUP, before GWAS, but with an identity matrix **I** fitted instead of the pedigree-based relationship matrix **A**) were used as phenotypes. Color version available online.

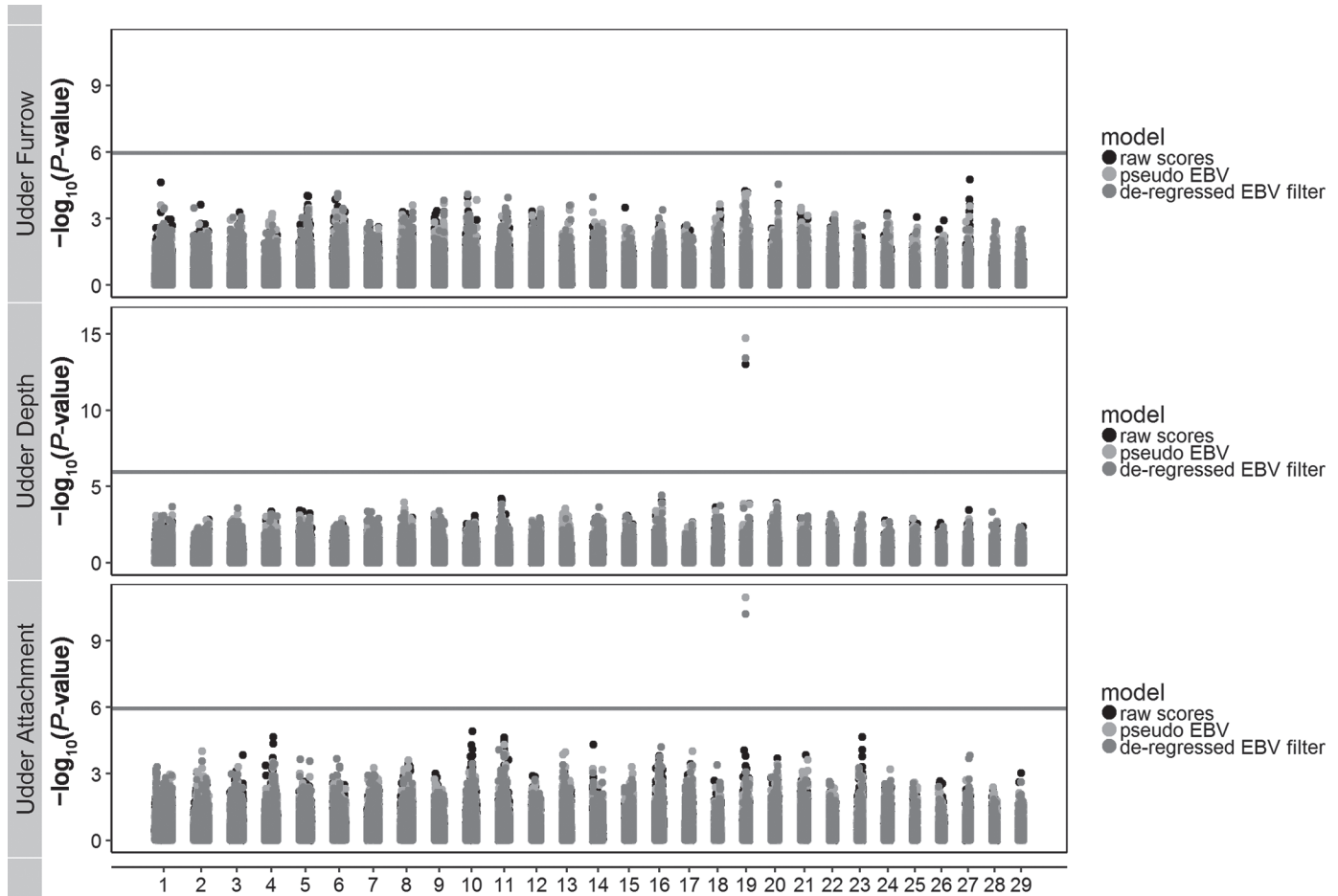


Figure 2. Manhattan plot of genome-wide association study (GWAS) for udder conformation traits (udder furrow, udder depth, udder attachment) using the Illumina Caprine 50K BeadChip (Illumina Inc., San Diego, CA) in dairy goats. P -values [$-\log_{10}(P\text{-values})$] for each SNP are shown on the y-axis, and each chromosome number is indicated on the x-axis. The solid (red) line indicates the threshold value for genome-wide significance. The legend indicates the phenotypes represented in the graph. Filtered deregressed EBV (EBV for animals that had their own conformation scores, or the reliability of their EBV was >0.6), raw scores, and pseudo-EBV (calculated by adjusting for fixed effects using BLUP, before GWAS, but with an identity matrix I fitted instead of the pedigree-based relationship matrix A) were used as phenotypes. Color version available online.

10.3168/jds.2017-12919). The proportion of the total variance explained by the significant SNP was between 0.4 to 7.0% (Supplemental Table S3).

Three genome-wide significant SNP were found for the analyzed conformation traits (Figure 2 and Supplemental Figure S4; <https://doi.org/10.3168/jds.2017-12919>). They were all located on chromosome 19, with the SNP snp23997-scaffold244-438286 being between exon 6 and 7 of *ASGR2* gene (*GOAT_ENSP00000347140*). The SNP snp24012-scaffold244-1259949 was located between exon 14 and 15 of the *ALOXE3* gene (*GOAT_ENSBTAP00000034797*). The snp24017-scaffold244-1460443 was 5,712 bp away from the nearest gene (Table 3). Additionally, 25 SNP were significant on a chromosome-wise level (Supplemental Table S3;

<https://doi.org/10.3168/jds.2017-12919>): 5 SNP for udder conformation, 9 for teat conformation (Supplemental Figure S5; <https://doi.org/10.3168/jds.2017-12919>), and 11 for feet and legs conformation. The SNP significant for the conformation traits explained between 0.1 and 13.8% of the total variance. It is worth noting that the lowest percentage of variance was attributed to the SNP significant for feet and legs conformation, particularly those that were significant for the psEBV model but had very low $-\log_{10}(P\text{-values})$ for the raw scores (i.e., snp12882-scaffold1498-142431). The highest percentage of variance explained was attributed to snp24012-scaffold244-1259949, which was genome-wide significant for udder depth with all of the analyzed models except of the deregressed EBV.

DISCUSSION

Milk Yield

The aim of the present study was to identify SNP significantly associated with milk yield and conformation traits in a commercial dairy goat herd. In total, 2 SNP associated with milk yield were found at the genome level, and a further 5 were significant at the chromosome level. Review of the available literature provides evidence for associations in dairy goats between milk yield and polymorphisms within *POU1F1* [*Capra hircus* autosome (**CHI**)7], a gene that is a positive regulator for growth hormone and prolactin (Lan et al., 2007); β -lactoglobulin gene (**CHI**11; Dettori et al., 2015; El Hanafy et al., 2015); *LALBA* (**CHI**5; Dettori et al., 2015); *MTHFR* (**CHI**6), a gene with known links to milk protein synthesis (An et al., 2015); *STAT5A* (**CHI**19; An et al., 2013); and *PRLR* (**CHI**20; Hou et al., 2013). However, these were not confirmed in the current study. Very few studies have used GWAS methodology to identify QTL in relation to milk yield in goats. Wasike et al. (2016) performed GWAS on a small sample ($n = 48$) of goats; however, the top 100 SNP found to be associated with milk yield accounted for only 3.1% of the genetic variance. Studies also suggest that the growth hormone gene, located on **CHI**19, could be associated with milk production in goats (Malveiro et al., 2001; Marques et al., 2003; Dettori et al., 2013). Current analysis identified a major QTL on **CHI**19 (at 26,066,457 bp) that was associated with milk yield; however, the significant SNP was not within the growth hormone gene (located on **CHI**19 between 47,266,042 and 47,267,410 bp) that has been reported to influence milk yield and composition. Martin et al. (2017) identified significant SNP for milk production traits on chromosome 19. The QTL for protein yield located at 28.5 Mb had a confidence interval (24.8–29.0 Mb) that overlaps with the SNP found in the current study at 26.1 Mb. The remaining QTL found by Martin et al. (2017) did not overlap with result of the current study.

None of the SNP significantly associated with milk yield in the present study were located within any of the gene regions previously associated with milk yield in dairy goats. As annotation of the goat genome is still in an early stage, it might be useful to utilize the homology between the bovine and caprine genomes, which has been well established in earlier studies (Ponce de Leon et al., 1996; Vaiman et al., 1996; Piumi et al., 1998; Dong et al., 2013).

The genome-wise significant SNP on chromosome 19 identified in the present study, which accounted for

between 6 and 7% of variance, is located approximately 15 kb downstream of the arachidonate 12-lipoxygenase (*ALOX12*) gene. The homologous bovine gene, positioned on chromosome 19, has been associated with several biological processes such as unsaturated fatty acid metabolic and biosynthetic processes, and has shown differential expression in dairy cows with extremely high and low milk yields (Bai et al., 2016; Yang et al., 2016). The genome-wise significant SNP located on chromosome 8 was located within a solute carrier family 1 (*SLC1A1*) gene, a trans-membrane transport channel. No studies in any livestock species have previously linked this gene to milk yield. Finally, the chromosome-wise significant SNP located on chromosome 14 is located within an uncharacterized transcript variant (LOC102186225), which is located between the genes *FAM49B* (approximately 1.5 Mb downstream) and *MYC* (approximately 100 kb upstream). The region encompassing these genes has been previously associated with major effect on milk yield, in a QTL study by Coppieters et al. (1998). Diacylglycerol acyltransferase 1 (*DGAT1*) has been shown to be strongly linked to milk yield in dairy cattle (Grisart et al., 2002; Winter et al., 2002) and milk fat in dairy goats (An et al., 2013). In goats, *DGAT1* is located on chromosome 14 (between 11,256,487 and 11,265,239 bp); however, the significant SNP located on chromosome 14 (at 23,135,282 bp) in the present study was not within the *DGAT1* gene. A recent study on French dairy goats conducted by Martin et al. (2017) identified SNP that were pointing toward *DGAT1* as the gene affecting fat content.

Conformation Traits

Conformation traits are of interest because of their associations with health and production. They are also very important for adaptation to machine milking. To our knowledge, there has been only one report on SNP associated with udder type traits (Maroteau et al., 2013). In case of feet and legs conformation, no significant SNP have been reported in goats. In the current study, genome-wise significant SNP were only found for udder attachment, udder depth, and front legs. It is worth noting that they were all in the same genomic region of chromosome 19. A SNP significantly associated with udder attachment was located between exons 6 and 7 of *ASGR2* gene. The homologous bovine gene is also located on chromosome 19 and is involved in regulation of protein stability and lipid homeostasis. Udder tissue is mainly composed of fat deposition and connective tissue (i.e., fibroblasts, adipocytes). Because both play an important role in udder formation, we

could hypothesize that processes affecting protein stability and especially lipid homeostasis may affect udder attachment. Moreover, a study conducted with microsatellite markers in cattle found a 1.6-Mb region covering the *ASGR2* gene that was significantly associated with udder attachment (Schrooten et al., 2000). Another genome-wise significant SNP was found for udder depth and was located between exons 14 and 15 of the *ALOXE3* gene, another member of the arachidonate lipoxygenase family. As well as the above-mentioned biological processes for ALOX12, both enzymes are involved in establishment of skin barrier; and ALOXE3 is involved in the processes of differentiation to fat cells, sphingolipid metabolism, and ceramide biosynthesis. Therefore, its possible involvement in udder conformation can be supported by its activity in fat metabolism, cell differentiation, and proliferation. The last genome-wise significant SNP was found for front legs with only the psEBV model. This SNP was located in a gene-rich area of chromosome 19. The nearest genes were *ARHGEF15* (rho guanine nucleotide exchange factor 15) and *ODF4* (outer dense fiber 4). The *ARHGEF15* gene encodes a protein annotated to be regulating stress fiber assembly, whereas *ODF4* encodes a protein localized in the outer dense fibers of tails of mature sperm cells. To date, this region has not been associated with any functions that could be directly linked to feet and legs conformation. Maroteau et al. (2013) also found SNP associated with udder type traits on chromosome 19. Unfortunately, due to the lack of SNP positions, it was impossible to compare genomic regions found in that study with the current results. In case of the remaining traits (udder furrow, teat shape, teat angle, teat placement, back legs, front feet, back feet), multiple chromosome-wise significant SNP were found. This seems to point toward a polygenic determinism of these traits.

Several studies in cattle and sheep have identified QTL for conformation traits (Schrooten et al., 2000; Ashwell et al., 2005; Cole et al., 2011; Gutiérrez-Gil et al., 2011; Wu et al., 2013). Initially, the studies were based on microsatellite markers and identified QTL for foot angle (BTA5, BTA6, BTA17, BTA21, BTA23), teat placement (BTA5, BTA6, BTA10, BTA19, BTA20, BTA21), udder depth (BTA5, BTA6, BTA13, BTA19, BTA27), teat length (BTA7, BTA13, BTA23), and rear leg set (BTA6, BTA13, BTA21, BTA24) (Gutiérrez-Gil et al., 2011; Lan et al., 2013; Wu et al., 2013). More recent studies involved SNP chips that cover the genome at much higher density. Significant SNP for feet and legs, foot angle, and rear leg side view in cattle have been reported on BTA2, BTA11, BTA13, BTA20, BTA26 (Cole et al., 2011; Wu et al., 2013).

Teat placement was significantly associated with SNP located on BTA6, BTA7, BTA11, and BTA19 (Wu et al., 2013), whereas significant SNP for teat length were reported on BTA11, BTA21, and BTA26 (Cole et al., 2011). Moreover, SNP associated with udder conformation were found on BTA2, BTA3, BTA5, BTA6, BTA7, BTA11, BTA12, BTA13, BTA14, BTA16, BTA17, BTA20, BTA22, BTA23, BTA25, and BTA26 (Cole et al., 2011; Sodeland et al., 2011; Flury et al., 2014). The current analysis provided no evidence as to the involvement of the above-mentioned genes in formation of conformation traits in the current goat population.

Further Discussion

Our aim was to perform an association analysis for conformation traits and milk yield using the caprine 50K SNP chip. Given the small sample size, the power to detect associations was low, which might have led to a situation where the detected effects were overestimated (Ball, 2013). This problem might be important in case of the conformation traits where the analyzed group included only 402 animals. It should be less apparent in the case of milk yield where the sample contained over 2,000 animals. Because of that, associations with SNP significant for conformation should be treated as an indication and certainly require further research to be validated. It should be also mentioned that annotation of the goat genome is still in a very early stage and, although bovine homologs can help identify gene regions to some extent, many regions remain unknown. With new and updated annotation, based on resequencing of the goat genome using single-molecule sequencing (Koren et al., 2016) from Pacific Biosciences (Menlo Park, CA) and BioNano Genomics (San Diego, CA), our ability to find candidate genes should improve.

Some concern might be raised as to the effect of breed composition on the results of the current GWAS analysis. This issue was addressed in our previous analyses on the implementation of genomic selection in the investigated population. Based on the principal component analysis as well as extent of linkage disequilibrium, we concluded that the investigated population is mostly homogeneous (Mucha et al., 2015); therefore, we have treated it as a new synthetic breed. Population structure should not have a strong negative effect on the power to detect QTL in the current study. On the other hand, the significant SNP identified in this study might not be segregating in other populations. This is mainly because linkage disequilibrium patterns are generally not consistent across breeds (de Roos et al., 2008), and may differ between divergently selected lines of the same breed (Banos and Coffey, 2010).

Deregressed EBV were used as pseudo-phenotypes to maximize the use of available information on animals. This method appears to be very effective provided that the reliability of the EBV is high (Garrick et al., 2009). Additionally, deregression was done with full pedigree, which removes parent average effect and makes deregressed EBV more suitable for GWAS (Ekine et al., 2014). In the case of milk yield, reliability of EBV was between 0.98 and 0.99 (1st and 3rd quantile), which should provide sufficiently accurate pseudo-phenotypes for GWAS. Conformation traits had a much lower reliability (1st and 3rd quantile between 0.07 and 0.32), which might lead to many false-positive results. Because of the nature of conformation traits, there was only a single measurement for each phenotyped animal. This, in combination with low heritabilities for these traits (0.02 to 0.38; McLaren et al., 2016), resulted in the low reliabilities observed. In contrast, moderate heritabilities (up to 0.45 in lactation 1; Mucha et al., 2014) and the repeated-records nature of milk yields resulted in high reliabilities.

Because deregressed EBV for conformation were not deemed accurate enough for GWAS, the use of raw conformation scores and psEBV were considered as alternative phenotypes. Of the 28 SNP found to be associated with the conformation traits, 2 were confirmed with all of the analyzed phenotypes (filtered deregressed EBV, psEBV, and raw scores). It is worth noting that the most significant SNP for conformation traits (udder depth) identified on chromosome 19 was confirmed with all 3 phenotypes. In case of raw scores and psEBV, only 4 of the 28 SNP were found to be in agreement. Additionally, several other SNP were found to be significant using deregressed EBV but these results were not confirmed with any of the other phenotypes. This might indicate that results obtained with deregressed EBV are likely to be false positives. Overall, results for udder and teat conformation traits were very similar across both phenotypes tested; however, there was a wider discrepancy between the psEBV and raw scores for feet and leg conformation traits. These traits may be better suited to genomic selection, to take full advantage of the small but significant genetic variance. In contrast, the largest single effect was found for udder depth, with a single SNP accounting for between 12.5 and 13.8% of variance. This trait also had the highest heritability (0.38). Pseudo-EBV were calculated using BLUP, which adjusted for fixed effects, without including any pedigree information. The weakness of adjusting for fixed effects before GWAS is that any error associated with fixed effects cannot be taken into account by the final model (Garrick and Fernando, 2013); therefore, where deregressed EBV show insufficient reliabilities, it

is preferable to simultaneously adjust for fixed effects within the GWAS analysis.

CONCLUSIONS

This study aimed to identify SNP associated with milk yield and conformation in dairy goats. The GWAS analyses resulted in 2 genome-wide significant SNP for milk yield located on chromosome 19, and 6 chromosome-wise significant SNP on chromosomes 4, 8, 14, 28, and 29. Moreover, 3 genome-wide significant SNP for conformation of udder attachment, udder depth, and front legs were identified on chromosome 19. These were accompanied by chromosome-wise significant SNP on chromosomes 4, 5, 6, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 21, 23, 27, and 28. Our results suggest that both milk yield and conformation traits have a polygenic background because, for most of them, we did not identify any QTL with major effects. We were unable to identify any candidate genes for milk yield or conformation.

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